# **Cerebrospinal fluid biomarkers for Alzheimer's**

eJIFCC. The Electronic Journal Of the International Federation Of Clinical Chemistry And Laboratory Medicine

## disease: their role in Clinical Chemistry

Kaj Blennow, MD, PhD<sup>1,2</sup> and Douglas Galasko, MD, PhD<sup>3</sup>

- Dept. of Clinical Neuroscience, Unit of Neurochemistry, University of Göteborg, Sweden
- 2 The Medical Research Council, Sweden.
- 3 Dept. of Neurology, VA Medical Center, Dept. Of Veterans Affairs, San Diego, California, USA

Corresponding author:

Kaj Blennow, MD, Ph.D.

Dept. of Clinical Neuroscience, Unit of Neurochemistry

Sahlgren's University Hospital, Mölndal

SE-431 80 Sweden Tel: + 46 31 3431791 Fax: + 43 31 3432426

E-mail: KAJ.BLENNOW@MS.SE

### In this article

 Key words: Alzheimer's disease (AD), β-amyloid (Aβ), biochemical markers, cerebrospinal fluid (CSF), diagnosis, tau.

### **SUMMARY**

### **ACKNOWLEDGEMENTS**

Supported by grants from the Swedish Medical Research Council (projects # 11560 and 12103).

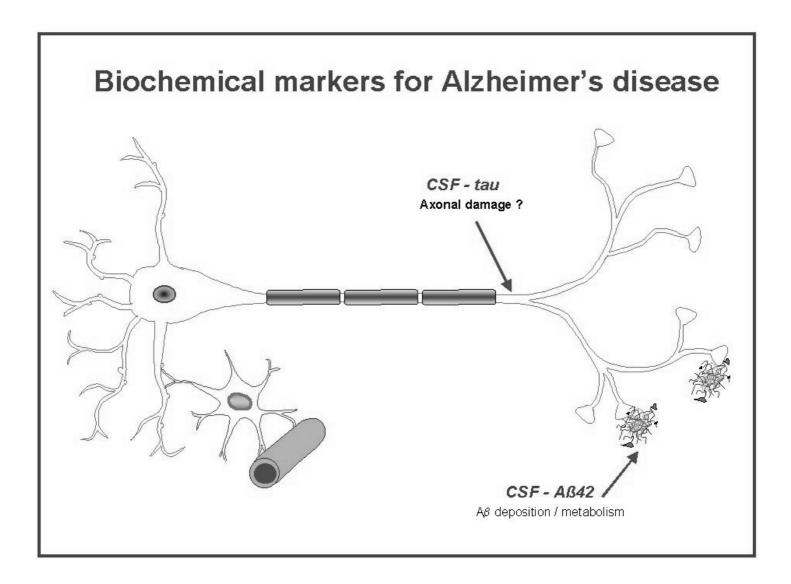
### **ABSTRACT**

In view of current (AChE inhibitors) and future (e.g. anti-Aβ aggregators), development and evaluation of cerebrospinal fluid (CSF) biomarkers for Alzheimer's disease (AD) has become a rapidly growing research field. Diagnostic biomarkers for AD would be especially valuable as aids in the diagnosis early in the course of the disease, when correct diagnosis is difficult, and when therapeutic compounds have the greatest potential of being effective. This paper reviews CSF biomarkers for AD, with emphasis on their role in the clinical diagnosis, and methodological aspects of importance for developing such analyses. Today, two biochemical markers, CSF-tau and CSF-Aβ42, perform satisfactory enough to have a role in the clinical workup of patients dementia, if used together with the cumulative information from clinical information and brain-imaging techniques. These markers are especially useful to discriminate early or incipient AD from age-associated memory impairment, depression, and some secondary dementias.

### INTRODUCTION

Alzheimer's disease (AD) is the major cause of dementia in the elderly. Although rare genetic (autosomal dominant) forms of AD exist, most patients have no obvious family history and are classified as having sporadic AD. The neuropathology of AD shows neuronal and synaptic degeneration, and an increased number of senile plaques (SP) and neurofibrillary tangles (NFT) compared to non-demented individuals of comparable age. Synaptic degeneration in AD is found in widespread cortical areas (Masliah et al, 1991). SP are composed of a central core of aggregated  $\beta$ -amyloid (A $\beta$ ) (Masters et al, 1985), a breakdown product derived from the amyloid precursor protein (APP) (Kang et al, 1987). NFT are insoluble intracellular thread-like structures made up of a hyperphosphorylated form of the microtubuleassociated protein tau, called phospho-tau (Goedert et al, 1993).

Today, several acetylcholine esterase inhibitors are available for symptomatic treatment of AD. Drugs that also may have beneficial effects on the disease process, e.g. compounds affecting the deposition of  $A\beta$ , are under development. These possibilities for therapeutic intervention have heightened awareness of the impor-



tance of early and accurate diagnosis of AD.

However, current clinical criteria for the diagnosis of AD are relatively vague, and are largely based on the exclusion of other dementing illnesses (McKhann et al, 1984). A relatively high accuracy rate with regard to the clinical diagnosis of AD (80-90%) has been reported (Tierney et al. 1988; Jellinger 1996; Galasko et al. 1994). However, these reports emanate from expert research academic centers and are often based on patients in the later stages of the disease who were followed for several years before the confirming autopsy. The diagnostic accuracy rate is probably considerably lower in general hospitals, and especially in the earlier stages of the disease when the symptoms are often silent or indistinct and clinical diagnosis is more difficult. This is unfortunate, as pharmaceutical therapy is probably most effective early in the course of disease, before neurodegeneration is too severe and widespread. Thus, there is a

great need for biochemical diagnostic markers (biomarkers) that could aid in the diagnosis of AD early in the course of the disease.

The cerebrospinal fluid (CSF) is in direct contact with the extracellular space of the brain, and its constituents reflect many biochemical changes in the brain. Since AD pathology is restricted to the brain, CSF is an obvious source of biomarkers for AD. During the last years, CSF biomarkers for AD have also gained increased attention. In this paper, we review the two CSF biomarkers that have been most extensively studied by different research ne ters and have proved to have the highest clinical diagnostic potential, i.e. CS Tau and CSF-AB42 (Fig. 1). We also focus on neurchemical factors of importance for making these biomarkers useful in clinical chemistry.

Besides their diagnostic potential, CSF biomarkers may also be useful to monitor the biochemical effect of therapeutic compounds. In AD, a drug that slows or arrests neurodegeneration might lower the level of a marker of active neuronal damage, such as CSF-tau. Similarly, a drug that acts by decreasing the deposition of  $A\beta$  might increase the level of CSF  $A\beta42$  in repeated samples.

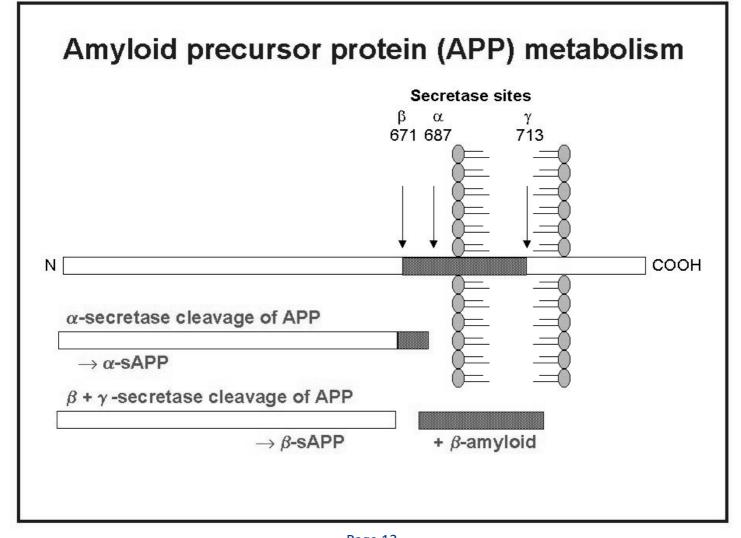
# CU RR EN T CSF biomarkers for an

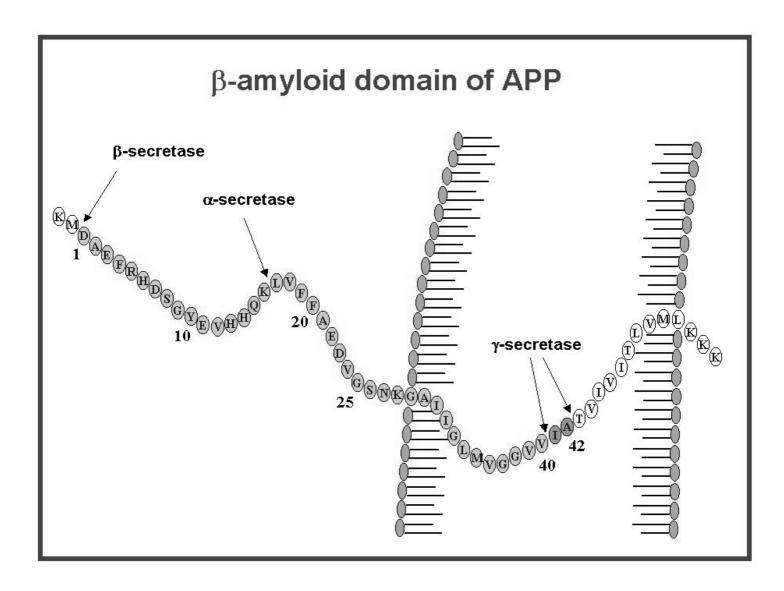
### CSF - (total) tau protein

Tau is a microtubule-associated protein located in the neuronal axons, while it is not present in dendrites (Goedert, 1993). There are six different isoforms, and numerous phosphorylation sites, of tau in the human brain (Goedert 1993).

An increase in CSF-(total)tau in AD has

been recorded in numerous studies (for review see e.g. Andreasen et al. 1998; Galasko 1998). The ability of CSF-tau to discriminate between AD and normal aging has been relatively good, above 80%, in most studies. High CSF-tau levels are, however, also found in a proportion of cases with other dementia disorders. This applies particularly to vascular dementia, in which high CSF-tau levels have been found in a relatively high proportion of cases in some studies (Blennow et al. 1995; Andreasen et al. 1998), while only in occasional cases in other studies (Tato et al. 1995; Mori et al. 1995; Arai et al. 1998; Mecocci et al. 1998; Nishimura et al. 1998; Hulstaert et al. 1999). It has been suggested that vascular dementia patients with high CSF-tau levels may constitute a subgroup with concomitant AD pathology (Andreasen et al. 1998), which implies that





CSF-tau might be of use in identifying vascular dementia cases where AD is a contributory factor to the dementia.

In contrast, in patients with other types of dementia (e.g. alcoholic dementia), chronic neurological disorders (e.g. Parkinson's disease, progressive supranuclear palsy) and psychiatric disorders (e.g. depression), elevated CSF-tau levels are found only in occasional cases (Blennow et al. 1995; Molina et al. 1997; Ellis et al. 1998; Mitani et al. 1998; Morikawa et al. 1999; Urakami et al. 1999).

The major clinical usefulness of CSF-tau seems to be in the discrimination of AD from normal aging. It might also be of use

in some other differential diagnoses (e.g. depression, alcoholic dementia and Parkinson's disease), which sometimes may be difficult to differentiate form AD on clinical grounds. The fact that an increase in CSF-tau can be found in acute destructive conditions, such as stroke (Arai et al, 1995), does not really reduce the clinical usefulness of CSF-tau, since these disorders are not differential diagnoses from AD

Most likely, the level of CSF-tau reflects the degree of neuronal/axonal degeneration, regardless of cause. Longitudinal data on CSF tau shows that levels remain stably elevated over 12–24 months of follow-up (Andreasen et al, 1999a). This would enable CSF-tau levels to be used as an outcome measure to assess treatment aimed at neuroprotection.

### CSF-AB42

The  $\beta$ -amyloid protein (A $\beta$  or  $\beta$ /A4 protein) is a cleavage-product from the amyloid precursor protein (APP), encoded by a single gene on chromosome 21 (Kang et al, 1987). APP is a transmembrane protein with a single transmembrane domain, a long N-terminal segment and a shorter cytoplasmic C-terminus (Fig 2). The AB part of APP encompasses the first 28 extracellular and the following 12-14 transmembrane amino acids (Fig 2). amyloidogenic secretory forms of APP is normally generated by cleavage within the Aβ region by an unidentified enzyme termed \alpha-secretase, while A\beta is produced by an alternative metabolic pathway, in which two proteases, termed \( \beta \)- and \( \gamma \)secretase cleave APP on each side of the Aβ sequence (Fig 2).

There are two major C-terminal variants of  $A\beta$ , a shorter form ending at Val-40 ( $A\beta$ 40), and a longer form ending at Ala-42 ( $A\beta$ 42) (Fig 3).  $A\beta$ 42 aggregates more rapidly than  $A\beta$ 40 and is also the predominating form of amyloid in diffuse plaques and in SP (for review see Dickson et al. 1997).

Aβ is generated as a soluble peptide during normal cellular metabolism, and is secreted into the extracellular space and biological fluids, including CSF (Haass et al. 1992). A marked decrease in CSF-AB42 is found in a high percentage of patients with AD (Motter et al. 1995; Galasko et al. 1998; Andreasen et al. 1999b; Hulstaert et al. 1999). The sensitivity is above 80-90%, resulting in a relatively good ability for CSF-Aβ42 to distinguish AD from normal aging and depression. However, the specificity for CSF-Aβ42 for the diagnosis of AD compared to other dementias has been less extensively studied than for CSF-tau, and needs to be further evaluated.

Measurement of AB in CSF may reflect cerebral amyloid deposition. Hypothetically, in AD AB secreted from neurons, and possibly other cells in the brain, binds to existing aggregates of AB in extracellular SP, with lower levels remaining to circulate in the CSF (Andreasen et al, 1999b). Longitudinal data show that CSF-AB42 levels remain relatively stable over 12–24 months of follow-up (Andreasen et al, 1999b). This implies that CSF could be used to monitor the effects of anti-amyloid drugs in AD.

# Co mb ination of CSF-tau and CSF-A642

The strategy of combining CSF-tau and CSF-A $\beta$ 42 as biomarkers for AD is appealing, since the concentrations of these substances are believed to reflect two of the central pathogenic processes in the disorder, and the combination might thus result in increased sensitivity and specificity. Indeed, some large studies have shown that both sensitivity and specificity increase for the combination compared with CSF-tau or CSF-A $\beta$ 42 alone (Galasko et al, 1998; Kanai et al, 1998; Hulstaert *et al.* 1999, Andreasen et al, 2000). These studies are summarized in Table 1.

Also when used as routine analyses in clinical chemistry, and the sensitivity and specificity figures are determined on all consecutive patients admitted for investigation of cognitive disturbances during one year in a community-based setting, the sensitivity to identify AD is above 90% (Andreasen et al, 2000).

Further, the combination of CSF-tau and CSF-A $\beta$ 42 also has a high sensitivity to predict progression to AD in patients with mild cognitive impairment (Andreasen et al, 1999c). This finding shows that these CSF markers show abnormal values very early in the disease process, already before the clinical dementia.

### CSF-ta u and CSF-A \( 42 \) in clini-

# Aβ42 AS BIOMARKERS FOR ALZHEIMER'S DISEASE. SENSITIVITY AND SPECIFICITY FIGURES FOR THE COMBINATION OF CSF-TAU AND CSF-

| Disorder   | =                          | Sensitivity | Specificity        | Comment  | Reference           |
|--|----------------------------|-------------|--------------------|--|---------------------|
| AD<br>Controls<br>OND including<br>non-AD dementia | 82<br>60<br>74             | 90%         | 80%                | <ul><li>Multi-center study in USA (6 centers)</li><li>Clinical diagnosis</li><li>Stored CSF samples.</li></ul>   | Galasko et al, 1998 |
| AD<br>Controls<br>OND including<br>non-AD dementia | 52<br>24<br>24<br>24<br>24 | 40%         | }90%               | <ul><li>Multi-center study in Japan (3 centers)</li><li>Clinical diagnosis</li><li>Stored CSF samples.</li></ul>   | Kanai et al, 1998   |
| AD<br>1999<br>Controls<br>OND<br>Non-AD dementia   | 150<br>100<br>84<br>79     | 85%         | 87%<br>86%<br>58%  | <ul> <li>nternational multi-center study (10 centers)</li> <li>Clinical diagnosis</li> <li>Stored CSF samples</li> <li>CSF analyses run at each research center</li> </ul> | Hulstaert et al,    |
| AD 2000 MICI Controls Depression Non-AD dementia   | 105<br>100<br>100<br>32    | 94%<br>75%  | 87%<br>100%<br>53% | <ul> <li>Prospective, community-based study</li> <li>Clinical diagnosis</li> <li>CSF samples run continuously in routine dinical neurochemistry.</li> </ul>                | Andreasen et al,    |
|  |                            |             |                    |  |                     |

### cal chemistry

As for all other test in clinical chemistry, methodological factors have to be evaluated when developing CSF biomarkers. CSF sampling, handling and storage can influence levels of many analytes. It has to be evaluated whether a molecule passes from serum to CSF across the blood-brain barrier, which occurs for many proteins that have higher levels in serum than in CSF (Tibblin et al, 1977). If so, the CSF levels will reflect the periphery more than the CNS. Neither CSF-tau nor CSF-AB42 are affected by this problem (Blennow et al, 1995; Vanderstichele et al, 1998). Further, CSF-tau or A\(\beta\)42 do not show concentration gradients in lumbar CSF (Vanderstichele et al, 1998), which, if present, complicates the interpretation of lumbar CSF levels, since these will vary with the volume and portion of CSF analyzed (Blennow et al, 1993).

However, the hydrophobic  $A\beta$  peptide may absorb to some types of test tubes commonly used for lumbar puncture or for centrifugation in the laboratory (Andreasen et al, 1999b). The level of  $A\beta42$  decreases to about 65% in polystyrene or in glass tubes, as compared with polypropylene tubes (Andreasen et al, 1999b). Therefore, polypropylene tubes must be used both for CSF sampling and for centrifugation and storage.

Both CSF-tau and CSF-A $\beta$ 42 are determined using sandwich ELISA techniques, using antibodies directed against different epitopes of the proteins, making the analyses very sensitive and specific. Current ELISA assays for CSF-tau use monoclonal antibodies that detect all isoforms of tau independent of phosphorylation, and thus measure the "total" CSF-tau level (Blennow et al, 1995), while A $\beta$  ELISA assays are specific to A $\beta$ 42, with minimal cross-reactivity against peptides ending at residues 43 or 40 (Motter *et al.* 1995; Vanderstichele *et al.* 1998).

The biological variation is low for both CSF-tau (Andreasen et al, 1998) and CSF-

A $\beta$ 42 (Andreasen et al, 1999b), i.e. very similar CSF levels are found when longitudinal CSF samples are analyzed from individual patients. Also the analytical variation is low, the coefficient of variance (CV) for internal control samples when CSF-tau and CSF-A $\beta$ 42 are run as a routine clinical neurochemical analyses during one year is approximately 10% (Andreasen et al, 2000).

Finally, it is also of importance to consider clinical confounding factors when evaluating CSF biomarkers for AD. One problematic issue is that studies are most often performed on clinically diagnosed patients, and data on neuropathologically confirmed cases are scarce. Although the positive predictive value for the clinical diagnosis of AD (i.e. the probability that AD is present when the criteria are met) has been relatively high, about 85%, the neg ative predictive value (i.e. the probability that AD is not present when the diagnostic criteria are not met) has been considerably lower (Tierney et al. 1988; Jellinger 1996; Galasko et al. 1994). This is especially troublesome for some of the non-AD dementias (e.g. vascular dementia and fronto-temporal dementia). In fact, neuropathological studies have found that a high proportion (40-80%) of clinically diagnosed patients with vascular dementia have notable concomitant AD pathology (Jellinger 1996; Kosunen et al. 1996). The fact that current clinical diagnostic criteria cannot be considered to be of 'gold standard' quality results in that it is difficult to get high sensitivity and specificity figures for CSF biomarkers. Further, even if they are asymptomatic, age-matched control subjects may harbor presymptomatic AD lesions in their brains (Tomlinson & Henderson 1976; Davies et al, 1988; Price & Morris 1999), which also reduces the specificity figures of CSF biomarkers for AD.

### THE CLINIC ALUSE OF CSF blomarkers for AD

Much effort has focused on finding a single neurochemical marker for AD. This may be elusive unless the marker is re-



IFCC Technical Secretariat, 30 Rue Lionnois, F-54000 Nancy, France

Phone: +33 3 83 35 26 16 Fax: +33 3 83 32 13 22 Email: Chantal.Thirion@ifccts.u-

"Providing leadership in clinical chemistry and clinical laboratory service".

lated to a pathogenic step that is unique to AD. For example, neuronal and synaptic degeneration is not only found in AD but also in most chronic degenerative disorders of the brain. Similarly, deposition of AB is not specific to AD, but is also found in normal aging, dementia pugilistica, Lewy body dementia, and after acute brain trauma, while deposition of PHF to in inclusions such as tangles may form in normal aging, dementia pugilistica, myotonic dystrophy, and other forms of tau are found in inclusions in progressive supranuclear palsy and fronto-temporal dementia(Davies et al, 1988; Roberts et al, 1994; Mc Kenzie et al, 1996). Thus, since the central neuropathological findings in AD are not specific for AD, it is unlikely that one single biochemical marker will absolutely discriminate between AD and other dementia disorders.

Instead, the combination of several CSF biochemical markers (e.g. CSF-tau, CSF-Aβ42 and possibly other like phospho-tau and  $\alpha/\beta$ -secretase cleaved APP) could be used in conjunction with other diagnostic methods. The overall accuracy of the clinical diagnosis of AD may increase if the diagnosis is based on cumulative information gained from the clinical examination, brain-imaging techniques (e.g. SPECT and MRT scans), and CSF biochemical markers. As an analogy, the clinical diagnosis of myocardial infarction is based on the combination of clinical symptomatology, electrocardiogram, and biochemical markers (e.g. creatine kinase). Today, the CSF markers tau and Aβ42, when used as an adjunct to clinical diagnosis, have the potential to help to differentiate AD from some problematic differential diagnoses, especially age-associated memory impairment, depressive pseudo-dementia, Parkinson's disease, progressive supranuclear palsy and alcoholic dementia.

### REFERENCES

Masliah E, Terry RD, Alford M, De-Teresa R, Hansen LA. Cortical and subcortical patterns of synaptophysinlike immunoreactivity in Alzheimer's disease. Am J Pathol 1991;138:235-246. Masters CL, Simms G, Weinman NA, Multhaup G, McDonald BL, Beyreuther K. Amyloid plaque core protein in Alzheimer's disease and Down syndrome. Proc Natl Acad Sci 1985;82:4245-4249.

Kang J, Lemaire HG, Unterbeck A, et al. The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor. Nature 1987;325:733-736.

Goedert M. Tau protein and the neurofibrillary pathology of Alzheimer's disease. TINS 1993;16:460-465.

McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of department of health and human services task force on Alzheimer's disease. Neurology 1984;34:939-944.

Tierney MC, Fisher RH, Lewis AJ, et al. The NINCDS-ADRDA Work Group criteria for the clinical diagnosis of probable Alzheimer's disease: a clinicopathologic study of 57 cases. Neurology 1988;38:359-64

Jellinger KA. Diagnostic accuracy of Alzheimer's disease: a clinicopathological study. Acta Neuropathol 1996;91:219-220.

Galasko D, Hansen LA, Katzman R, Wiederholt W, Masliah E, Terry R, Hill LR, Lessin P, Thal LJ. Clinical-neuropathological correlations in Alzheimer's disease and related dementias. Arch Neurol 1994;51:888-95.

Andreasen N, Vanmechelen E, Van de Voorde A, et al. Cerebrospinal fluid tau protein as a biochemical marker for Alzheimer's disease: a community based follow up study. J Neurol Neurosurg Psychiatry 1998;64:298-305.

Galasko D. Cerebrospinal fluid levels of Aβ42 and tau: potential markers of Alzheimer's disease. J Neural Transm 1998 (suppl);53:209-221.



IFCC Technical Secretariat, 30 Rue Lionnois, F-54000 Nancy, France

Phone: +33 3 83 35 26 16 Fax: +33 3 83 32 13 22 Email: Chantal.Thirion@ifccts.u-

"Providing leadership in clinical chemistry and clinical laboratory service".

Blennow K., Wallin A., Ågren H., Spenger C., Siegfried J., Vanmechelen E. Tau protein in cerebrospinal fluid: a biochemical diagnostic marker for axonal degeneration in Alzheimer's disease? Mol Chem Neuropathology 26:231-245;1995.

Tato RE, Frank A, Hernanz A. Tau protein concentrations in cerebrospinal fluid of patients with dementia of the Alzheimer type. J Neurol Neurosurg Psychiatry 1995;59:280-283.

Mori H, Hosoda K, Matsubara E, et al. Tau in cerebrospinal fluids: establishment of the sandwich ELISA with antibody specific to the repeat sequence in tau. Neurosci Lett 1995;186:181-183.

Arai H, Satoh-Nakagawa T, Higuchi M, et al. No increase in cerebrospinal fluid tau protein levels in patients with vascular dementia. Neurosci Lett 1998;256:174-6.

Mecocci P, Cherubini A, Bregnocchi M, et al. Tau protein in cerebrospinal fluid: a new diagnostic and prognostic marker in Alzheimer disease? Alzheimer Dis Assoc Disord 1998;12:211-4.

Nishimura T, Takeda M, Nakamura Y, et al. Basic and clinical studies on the measurement of tau protein in cerebrospinal fluid as a biological marker for Alzheimer's disease and related disorders: multicenter study in Japan. Methods Find Exp Clin Pharmacol 1998;20:227-35.

Hulstaert F, Blennow K, Ivanoiu A, et al. Improved discrimination of AD patients using  $\beta$ -amyloid<sub>(1.42)</sub> and tau levels in CSF. Neurology 1999;52:1555-1562.

Molina JA, Benito-Leon J, Jimenez-Jimenez FJ, et al. Tau protein concentrations in cerebrospinal fluid of non-demented Parkinson's disease patients. Neurosci Lett 1997;238:139-41.

Ellis RJ, Seubert P, Motter R, et al. Cerebrospinal fluid tau protein is not elevated in HIV-associated neurologic disease in humans. HIV Neurobehavioral Research Center Group. Neurosci Lett 1998;254:1-

4.

Mitani K, Furiya Y, Uchihara T, Ishii K, Yamanouchi H, Mizusawa H, Mori H. Increased CSF tau protein in corticobasal degeneration. J Neurol 1998;245:44-6.

Morikawa Y, Arai H, Matsushita S, et al.

Cerebrospinal fluid tau protein levels in demented and nondemented alcoholics.

Alcohol Clin Exp Res 1999;23:575-7.

30 Rue Lioppois

Urakami K, Mori M, Wada K, et al. A comparison of tau protein in cerebrospinal fluid between corticobasal degeneration and progressive supranuclear palsy. Neurosci Lett 1999;259:127-9.

Arai H, Terajima M, Miura M, et al. Tau in cerebrospinal fluid: a potential diagnostic marker in Alzheimer's disease. Ann Service". Neurol 1995;38:649-652.

Andreasen N, Minthon L, Clarberg A, Davidsson P, Gottfries J, Vanmechelen E, Vanderstichele H, Winblad B, Blennow K. Sensitivity, specificity and stability of CSF-tau in AD in a community-based patient sample. Neurology 1999a;53:1488-1494.

Dickson DW. The pathogenesis of senile plaques. J Neuropathol Exp Neurol 1997;56:321-339.

Haass C, Schlossmacher MG, Hung AY, et al. Amyloid  $\beta$ -peptide is produced by cultured cells during normal metabolism. Nature 1992;359:322-325.

Motter R, Vigo-Pelfrey C, Kholodenko D, et al. Reduction of  $\beta$ -amyloid peptide42 in the cerebrospinal fluid of patients with Alzheimer's disease. Annals of Neurology 38:643-648;1995.

Galasko D, Chang L, Motter R, et al. High cerebrospinal fluid tau and low amyloid beta42 levels in the clinical diagnosis of Alzheimer disease and relation to apolipoprotein E genotype. Arch Neurol 1998;55:937-45.

Andreasen N, Hesse C, Davidsson P, Wal-



IFCC Technical Secretariat 30 Rue Lionnois, F-54000 Nancy, France

Phone: +33 3 83 35 26 16 Fax: +33 3 83 32 13 22 Email: Chantal.Thirion@ifccts.u-

"Providing leadership in clinical chemistry and clinical laboratory service".

lin A, Minthon L, Winblad B, Vanderstichele H, Vanmechelen E, Blennow K. Cerebrospinal fluid  $\beta$ -amyloid<sub>(1.42)</sub> in Alzheimer's disease: differences between early- and late-onset Alzheimer disease and stability during the course of disease. Arch Neurol 1999b;56:673-680.

Kanai M, Matsubara E, Isoe K, Urakami K, Nakashima K, Arai H, Sasaki H, Abe K, Iwatsubo T, Kosaka T, Watanabe M, Tomidokoro Y, Shizuka M, Mizushima K, Nakamura T, Igeta Y, Ikeda Y, Amari M, Kawarabayashi T, Ishiguro K, Harigaya Y, Wakabayashi K, Okamoto K, Hirai S, Shoji M. Longitudi na 1 study of cerebrospinal fluid levels of tau, A beta1-40, and A beta1-42(43) in Alzheimer's disease: a study in Japan. Ann Neurol 1998;44:17-26.

Andreasen N, Minthon L, Davidsson P, Vanmechelen E, Vanderstichele H, Winblad B, Blennow K. CSF-tau and CSF- $A\beta42$  as diagnostic markers for Alzheimer's disease in clinical practice. Submitted, 2000.

Andreasen N, Minthon L, Vanmechelen E, Vanderstichele H, Davidsson P, Winblad B, Blennow K. CSF-tau and CSF-Aβ42 as predictors of development of Alzheimer's disease in patients with mild cognitive impairment. Neurosci Lett 1999c;273:5-8.

Tibblin G, Link H, Öhman S. Principles of albumin and IgG analyses in neurological disorders. I. Establishment of reference values. Scand J Clin Lab Invest 1977:37; 385-390.

Vanderstichele H, Blennow K, D'Heuvaert ND, et al. Development of a specific diagnostic test for measurement of  $\beta$ -amyloid<sub>(1-42)</sub> in CSF. Progress in Alzheimer's and Parkinson's Diseases. Eds.: Fisher A, Hanin I, Yoshida M. Plenum Press, New York, 1998:773-778.

Blennow K, Fredman P, Wallin A, Gottfries CG, Långström L, Svennerholm L. Protein analyses in cerebrospinal fluid: I. Influence of concentration gradients for proteins on cerebrospinal fluid/serum al-

bumin ratio. Eur Neurol 33:126-128; 1993.

Kosunen O, Soininen H, Paljärvi L, Heinonen O, Talasniemi S, Riekkinen PJ Sr. Diagnostic accuracy of Alzheimer's disease: a neuropathological study. Acta Neuropathol 1996;91:185-193.

Tomlinson BE, Henderson G. Some quantitative cerebral findings in normal and demented old people, in Terry RD, Gershon S (eds.): Neurobiology of aging. New York, Raven Press, 1976:183-204.

Davies L, Wolska B, Hilbich C, et al. A4 amyloid protein deposition and the diagnosis of Alzheimer's disease. Neurology 1988;38:1688-1693.

Price JL, Morris JC. Tangles and plaques in nondemented aging and "preclinical" Alzheimer's disease. An n Neurol 1999;45:358-368.

Davies L, Wolska B, Hilbich C, et al. A4 amyloid protein deposition and the diagnosis of Alzheimer's disease. Neurology 1988;38:1688-1693.

Roberts GW, Gentleman SM, Lynch A., Murray L., Landon M., Graham DI. Beta amyloid protein deposition in the brain after severe head injury: implications for the pathogenesis of Alzheimer's disease. J Neurol Neurosurg Psychiatry 57;419-25:1994.

McKenzie JE, Edwards RJ, Gentleman SM, et al. A quantitative comparison of plaque types in Alzheimer's disease and senile dementia of the Lewy body type. Acta Neuropathol (Berl) 91;526-9:1996.



IFCC Technical Secretariat, 30 Rue Lionnois, F-54000 Nancy, France

Phone: +33 3 83 35 26 16 Fax: +33 3 83 32 13 22 Email: Chantal.Thirion@ifccts.u-

"Providing leadership in clinical chemistry and clinical laboratory service".